

SYNTHESIS, PHYSICOCHEMICAL ELUCIDATION AND BIOLOGICAL SCREENING STUDY OF NEW LIGAND DERIVED FROM 5,6-O-ISO PROPYLIDENE-L-ASCORBIC ACID AND ITS METAL(II) COMPLEXES

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Abstract: A simple chemistry method approach was used to synthesise new ligand derivate from L-ascorbic acid and its complexes. All of them were water-soluble and are used quite extensively in the medical and pharmaceutical fields. This study synthesised the new ligand derivative from L-ascorbic acid-base using the following steps: A 5,6-O-isopropylidene-L-ascorbic acid was prepared by reacting dry acetone with L-ascorbic acid followed by reacting it with trichloroacetic acid to yield [chloro(carboxylic)methylidene]-5,6-O-isopropylidene-L-ascorbic acid in the second stage. In the third stage, the derivative was reacted with (methyl(6-methyl-2-pyridylmethyl)amine) to create a new ligand (ONMILA). This novel ligand was identified using a number of techniques, namely mass spectroscopy, ¹H, ¹³C-Nuclear magnetic resonance, Fourier Transform Infrared (FT-IR), and Ultraviolet-visible (UV-Vis) spectra. It was observed that several complexes formed between the ligand and divalent metal ions (Co, Ni, Cu, Zn, Cd). Based on Micro Elemental Analysis, the mole ratio was (1:1) (M:L). Magnetic susceptibility, elemental analysis (C.H.N.O) procedures, molar conductivity tests, and proportion of metal ions calculations were used to describe the complexes. The findings showed the novel ligand had a mono, negative charge and behaved like a tridentate ligand type (N.N.O.). Therefore, the octahedral formula is suggested for all compounds. Only one spot was observed on thin layer chromatography (T.L.C.) for ligand (L) and complexes, indicating that the reaction completed and delivered only a single product. These chemicals have been connected to both Gram-negative and Gram-positive bacteria. The results suggested that antibacterial activity in metal complexes is higher than in the free ligand.

Keywords: L-ascorbic acid, ligand, techniques, metal complexes, antibacterial activity

1. Introduction

L-enantiomer of ascorbic acid without scurvy is a condition caused by vitamin C deficiency (Zümreoglu-Karan, 2006). The human body requires a robust immune system and therefore, a vitamin-rich daily diet is recommended. Vitamins with critical biological properties are abundant in natural foods, particularly vegetables and fruits, which are rich in cinnamon, vitamin C, and hesperidin and well-known for their health benefits (Bellavite & Donzelli 2020). L-ascorbic acid is a type of vitamin C which has water-soluble antioxidants and a protective function. Although most mammals can produce ascorbate, they cannot produce vitamin C (Chatterjee, 1973). L-ascorbic acid is a biological antioxidant (Loke et al., 2006) that protects the cell from damaging radicals, especially those generated during incomplete O₂ oxidation (Padayatty et al., 2003). In both chemistry and biology, L-ascorbic acid is a critical chemical and its complexes are significant in both (Hanukoglu, 2006). Although ascorbic acid has

many antimicrobial effects, some of its oxidative products are toxic (Hollis et al., 1985). According to Fodor (1983), L-ascorbic acid molecule has four hydroxyl groups which participate in conventional esterification. It is possible to use L-ascorbic acid (L-asc.) or Vitamin C and its derivatives in various biological and industrial uses (John & James, 2009). It has several donors, atoms, O(1), O(2), O(3), O(5), and O(6), that are usually involved in metal-ligand bonding (Tajmir-Riahi, 1990). The functional groups -OH, -COOH, -SH, -NH₂ in derivatives of L-asc. have been found to coordinate with metal ions and form complexes [10]. These vitamin-metal complexes are essential in development of medication and nutrition (Wasi, 1987) while metal complexes are crucial for the pharmaceutical and agricultural industries. A living system relies heavily on trace amounts of metal elements and these transition metal ions ensure that different enzymes function correctly (Hariprasath et al., 2010). The current study examined these metal complexes for their antiseptic action alongside various bacterial strains. Their antibacterial activity was found to be effective against all the strains. The study also attempted to establish and validate a new apparent synthesis and description of an L-ascorbic acid derivative ligand (L).

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2. Experimental

2.1 Instrumentation

Shimadzu FT-IR spectrophotometer ranging 4000-400 cm^{-1} , Shimadzu UV-Vis. spectrophotometer with 200-1100 nm^{-1} , Stuart electrothermal equipment, model SMP30 W.T.W. in OLAP cond 720 digital conductivity meter, Bruker D.R.X. (500-MHz, DMSO-d_6) spectrometer, mass spectrometer 5975 quadrupole. Elemental analyser Euro Vectro-3000A, Atomic absorption spectrophotometer Analytic Jena (A.A350), Magnetic Susceptibility Balance Mode (MSB-MKI). 854 Schwach Autoclave, Gallen Kamp Sterilizer, Memmert Incubator.

2.2 Materials

L-ascorbic acid, Trichloro acetic acid, Potassium hydroxide, Methyl(6-methyl-2-pyridylmethyl) amine, Metal chloride " $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$, $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$, $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$, ZnCl_2 , $\text{CdCl}_2 \cdot \text{H}_2\text{O}$ ", Mueller Hinton agar, dry acetone, N-Hexane, Dimethylsulfoxide, Dimethylformamide, HCl gas, Ethanol, Distilled water. "All chemicals used were of the highest purity. B.D.H., Fluka, and Merck were used without further purification.

2.3 Synthesis of The New Ligand (L)

2.3.1 Synthesis of 5,6-O-iso propylidene-l-ascorbic acid:

A 250 mL flask containing 0.528 grams (0.003 moles) of L-ascorbic acid powder along with 50 mL of dry distillers acetone was given a rapid bubble bath of dry hydrogen chloride and stirred for 20 minutes. The supernate was decanted after 20 mL of n-hexane was added followed by additional stirring and then chilling (ice-water). After each addition, the precipitate was rinsed with (100 mL) of the acetone-hexane combination (3:7) (v/v) followed by stirring, cooling in cold water, and then extracting the supernate. This process was repeated four times. The final precipitate was subjected to drying under decreased pressure, which resulted in the production of a white crystalline residue with a melting temperature of 216 $^{\circ}\text{C}$. This yielded 70.51 grams R_f (0.69) in benzene:ethanol (8:2) (v/v) solvent solution.

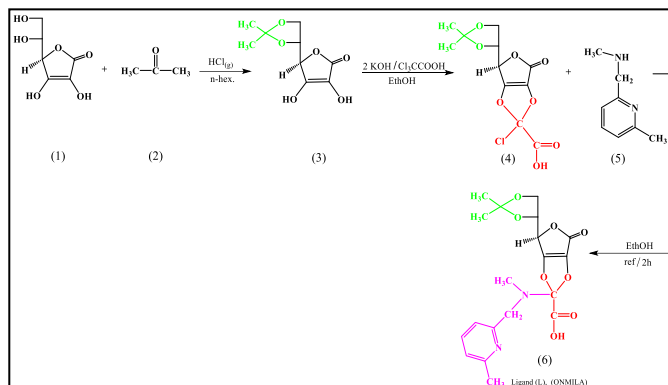
2.3.2 Synthesis of [Chloro(Carboxylic)Methylidene]-5,6-Isopropylidene-L-Ascorbic Acid:

5,6-O-isopropylidene-L-ascorbic acid (0.432 g, 0.002 mole) was dissolved in (25 mL) ethanol. A potassium hydroxide solution (0.112 g, 0.002 moles) in ethanol (20 mL) was later added and stirred for 30 minutes. Next, trichloro acetic acid (0.163 g, 0.001 mole) in ethanol (15 mL) solution was added and left for one hour. The product was filtered. Recrystallisation from 25mL (20 mL ethanol + 5 mL water) yielded 73.5% of pale brown solid crystalline residue with a melting point of 209 $^{\circ}\text{C}$. (benzene: methanol) R_f 0.57 in (5:5).

2.3.3 Synthesis of The Ligand (L), [O, O-2,3-(N-Carboxylicmethylidene)-N-Methyl-1-(6-Methylpyridyl]-5,6-Isopropylidene-L-Ascorbic Acid:

0.306 g (1 mmole) of methylidene-(chloro(carboxylic)-5,6-iso propylidene-L-ascorbic acid was diluted in 20 mL ethanol. Using methyl (6-methyl-2-pyridylmethyl)amine, the solution was

dropped wide (0.14 mL, 0.001 mole) and refluxed for two hours. A deep dark precipitate mass formed after some time at room temperature, and which was subsequently separated from the solution. Recrystallising the resulting mass in ethanol yielded a brown crystalline substance with a melting point of (127 $^{\circ}\text{C}$) and a yield of 80.47 percent. R_f (0.48) in a solvent solution of benzene and ethanol (8:2) (v/v). The diagram below (Scheme 1) describes the reaction.



Scheme 1. Synthesis route of a new ligand (ONMILA) derivative.

2.3.4 Synthesis of Metal Complexes

1 mmol metal chloride 0.238 g $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$, 0.237 g $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$, 0.170 g $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$, 0.136 g ZnCl_2 , 0.183 g $\text{CdCl}_2 \cdot \text{H}_2\text{O}$ in 10 mL ethanol solution was added to a mixture of ligand (0.406 g, 1 mmol) in 10 mL ethanol. After agitating the solutions for an hour, the complexes were gently precipitated by evaporation. Figure 9 describes the process of recrystallising the complexes from heated ethanol. Water, methanol, ethanol dimethylsulfoxide, and dimethylformamide are all effective for dissolving isolated complexes which are colourful solids that are stable in air but insoluble in normal organic solvents. Table 1 describes the physical characteristics and provides analytical data for ONMILA and its complexes.

2.4 Biological Activity

The agar-well diffusion technique was applied in the testing of synthesised ligand and complexes against *Staphylococcus aureus* and *Streptococcus pyogenes* which are Gram (+), *E-coli* and *Klebsiella pneumonia*, Gram (-). All of the compounds were found to be effective against the bacteria (Al-Khafagy, 2016; Valarmathy et al., 2020). Muller Hinton Agar was used as the medium for the cultivation of the microorganisms to be tested. Dimethyl sulfoxide (DMSO), was used as a solvent in a single concentration of 1×10^{-3} M to make the chemical solutions; this was aimed at studying its biological activity. The plates were kept in an incubator at 37 $^{\circ}\text{C}$ for a whole day. The compounds' antibacterial activity was evaluated with regard to the size of the inhibition zone they produced against the particular strain of bacteria tested. The growth inhibition zone for each sample was determined by utilising the mean value obtained from three separate repetitions in order to reach a conclusion.

3. Results and Discussion

3.1 Physical Characteristics and Elemental Examination

Table 1 shows physical features and results of elemental analysis (C.H.N.O) examination and the organized compounds' metal substances.

Table 1. Physical properties and analytical statistics of ONMILA and its complexes

Empirical formula	Colour	M.wt	M.P °C	Yield %	R _f	Element Analysis				Found (calcul.) Metal %
						Found (calcul.) %				
						C	H	N	O	
ONMILA= C ₁₉ H ₂₂ O ₈ N ₂	Deep brown	406.38	127	80.47	0.48	56.15 (55.93)	5.45 (5.31)	6.89 (6.94)	31.49 (31.7)	—
[Co(L)Cl ₂ (H ₂ O)]H ₂ O	Reddish-Brown	572.25	151	74.71	0.43	39.87 (39.49)	4.57 (4.59)	4.88 (4.67)	27.95 (27.81)	10.29 (9.74)
[Ni(L)Cl ₂ (H ₂ O)]3H ₂ O	Olive	608.04	145	70.13	0.41	37.53 (37.46)	4.97 (4.9)	4.6 (4.72)	31.57 (31.74)	9.65 (9.05)
[Cu(L)Cl ₂ (H ₂ O)]H ₂ O	Deep green	576.86	187	82.36	0.37	39.55 (39.71)	4.54 (4.42)	4.85 (4.54)	27.73 (27.9)	11.01 (11.24)
[Zn(L)Cl(H ₂ O) ₂]Cl.3H ₂ O	ale brown	632.74	191 °D	73.2	0.40	36.06 (35.45)	5.09 (5.23)	4.42 (4.69)	29.43 (29.49)	10.33 (10.17)
[Cd(L)Cl(H ₂ O) ₂]Cl.2H ₂ O	Deep beige	661.76	173 °D	65.68	0.35	34.48 (33.9)	4.56 (4.47)	4.23 (4.29)	29.01 (29.25)	16.98 (16.42)

3.2 N.M.R Spectra for the ONMILA Ligand

¹H-NMR spectrum in Figure 1 (500 MHz in DMSO-d₆) showed numerous signals. A carboxylic-OH low signal strength was seen at δ 11.85 ppm (s, 1H). The Pyridine ring showed multiplet signals at δ 7.7-7.21 (m, 3H) (Lee & McCarthy, 2019). A doublet peak at δ 5.25, δ 5.22 (d, 1H) was due to CH-4 lactone ring. The signals at δ 4.84, δ 4.73 (m, 2H) were assigned to CH-5, and the signals at δ 3.79 (s, 2H) and δ 3.71 (t, 2H) were attributed to pyridine-CH₂ and CH₂-6 l-ascorbic acid moieties, while that at δ 2.73 (s, 3H), 2.45 (s, 3H) were due to CH₃- pyridine and CH₃-N of 2-aminomethyl moieties. A single peak at δ 1.51 (s, 6H) ppm was attributed to (CH₃)₂ of 5,6-O-iso propylidene (Al-Noor et al., 2021).

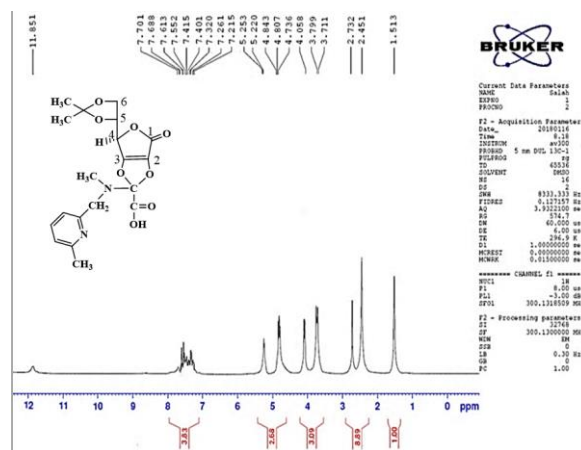


Figure 1. ¹H-NMR spectrum of ONMILA.

There was a carboxylic acid peak at 176.04 ppm in the ¹³C-NMR spectrum as shown in Figure 2, with lactones ring carbon and pyridine (C=N) signal also present. Carbons (C-2 and C-3) were responsible for the two peaks at (137.22 and 142.71) ppm. There is a possibility that the "double bond that was conjugated from (C-1 to C-3)" shifted the (C-3) signal upfield (Lee & McCarthy, 2019) while the signals in the range of (136,26-120.6) "ppm were dispensed to C=C" pyridine ring carbon atoms. The signals at (66.03) due to C10-N of 2-methylpyridine, and the peak at (55.95) were assigned to C9-N amino-methyl moieties, while the signals in the range of (73.6-66.03) ppm were assigned to (C5-C4) and (C5-C-6). C-C carbon atoms C-7 and C-8 were responsible for the signals at 33.81, 29.23, and 27.13 ppm respectively (Lee & McCarthy, 2019; Al-Noor et al., 2021).

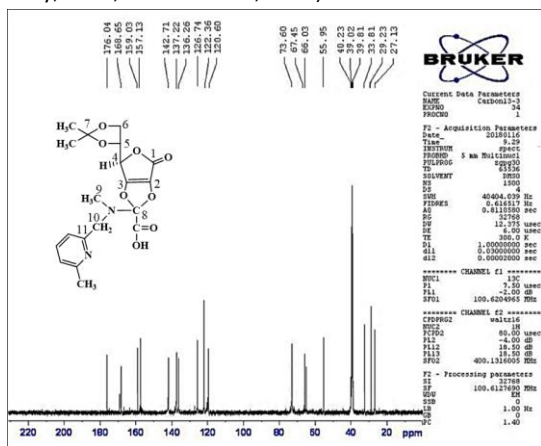


Figure 2. ¹³C-NMR spectrum of ONMILA.

3.3 Mass Spectrum for ONMILA

It was discovered that the molecular ion top at (m/z = 406.2) in the ligand's mass spectrum as described in Figure 3 corresponded with (C₁₉H₂₂N₂O₈) (Ahmed et al., 2015; Gao, (2021). Scheme 2 summarises the pieces and their relative abundance.

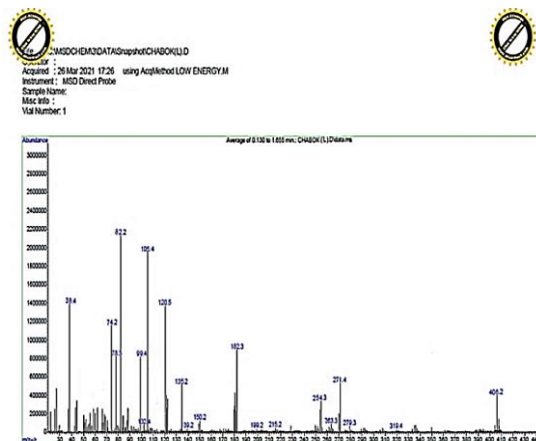
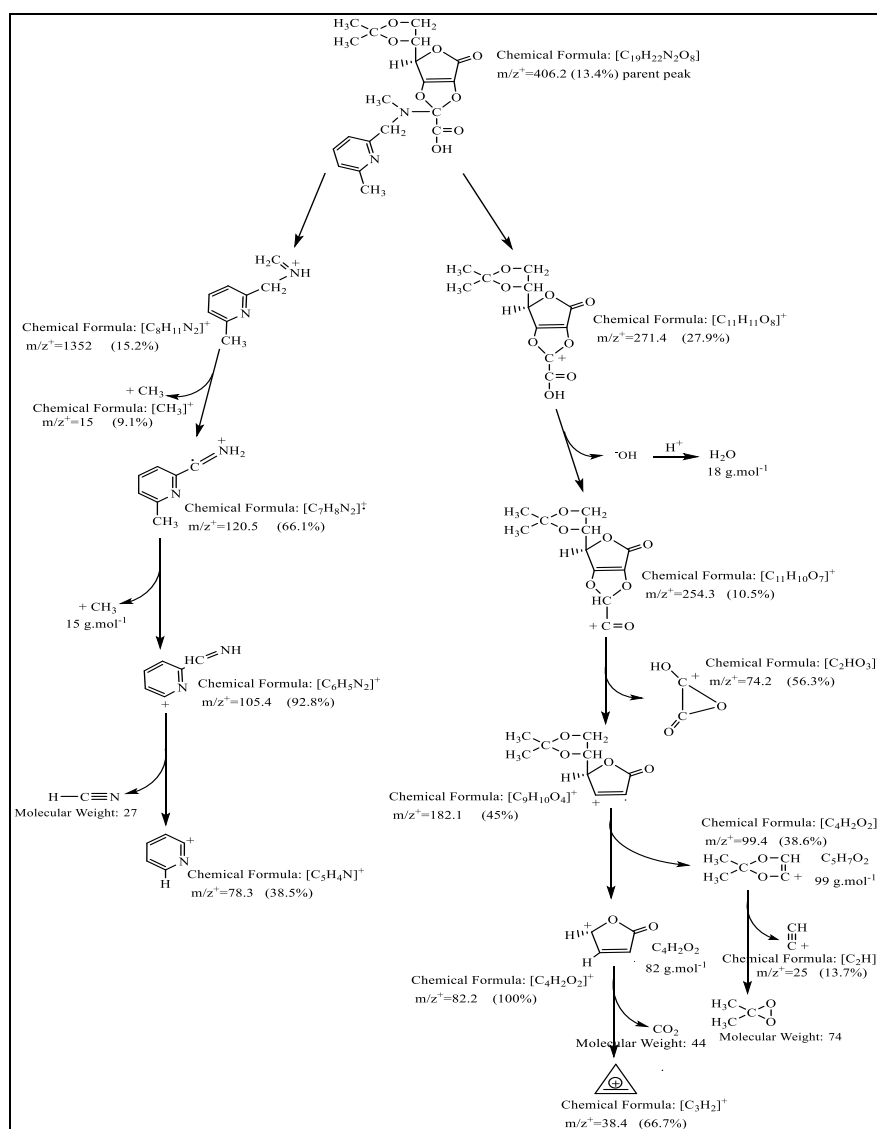


Figure 3. Mass spectrum of ONMILA.



Scheme 2. The fragmentation of ONMILA.

3.4 Infrared Spectra of ONMILA and its Complexes

A comparison of ONMILA (FT-IR) spectrum and its metal complexes (nickel complex, for example) is shown in Figure 4 and Figure 5. Table 2 summarises the major I.R. bands and their potential assignments. The frequency of the essential wide band at 3429 cm^{-1} , which may be attributed to the OH-carboxylic acid in the free ligand, did not change for any complexes in the range of $3435\text{--}3416\text{ cm}^{-1}$ (Waddai et al., 2015). A band that was produced at 1627 cm^{-1} and was ascribed to the (C=N) stretching vibration of pyridine-N was linked with $\nu(C=N)$ altered in shape and moved to a lower frequency in all complexes in the range of $1611\text{--}1595\text{ cm}^{-1}$ (El-Sonbati et al., 2016; Naji, 2014). Broad bands at $3419, 3435, 3416, 3432,$ and 3422 cm^{-1} due to (O-H) stretching in conjunction with OH-carboxyl stretching and bands

at $(807, 861, 855, 869,$ and $874)\text{ cm}^{-1}$ in Co(II), Cu(II), Ni(II), Zn(II), and Cd(II) complexes respectively attributable to coordination water were found in all complexes (Waddai et al., 2015; Al-Farhan et al., 2021). The stretching vibration to the (C=O) ring of lactones was found at $(1710)\text{ cm}^{-1}$, with a slight frequency shift in the range $(1695\text{--}1723)\text{ cm}^{-1}$ due to it not being involved in the coordination (Waddai et al., 2015; Radisavljević & Petrović, 2020). The stretching band (C=O)-carboxyl was found at $(1685)\text{ cm}^{-1}$ with a shape change and lower frequency in the range $(1668\text{--}1653)\text{ cm}^{-1}$ caused by a complex framework with the metal ions (Lawal et al., 2017). New weak bands displayed in all metal complexes spectra in the low-frequency range at $(518\text{--}483)\text{ cm}^{-1}$ and $(477\text{--}446)\text{ cm}^{-1}$ proved to bond (Kareem et al.; Raman & Sobha, 2010).

Table 2. Characteristics of FT-IR absorption bands of ONMILA and its complexes.

Empirical formula	ν OH Carbox., Hydrat.	ν C=O lactone	ν C=O Carbox.	ν C=N ν C=C	ν C-N pyridine	δ H ₂ O aqua	ν M-N	ν M-O
(L) ONMILA	3429,b	1710,m	1685,sh	1627,s	1155,m	-	-	-
[Co(L)Cl ₂ (H ₂ O)]H ₂ O	3419,b	1719,m	1653,s	1598,s	1118,m	807,w	518,w	454,w
[Ni(L)Cl ₂ (H ₂ O)]3H ₂ O	3435,b	1698,s	1668,m	1605,m	1125,w	861,w	483,w	446,w
[Cu(L)Cl ₂ (H ₂ O)]H ₂ O	3416,b	1714,m	1659,s	1611,s	1132,m	855,w	511,w	477,w
[Zn(L)Cl(H ₂ O) ₂]Cl.3H ₂ O	3432,b	1723,s	1666,s	1609,m	1121,w	869,w	487,w	461,w
[Cd(L)Cl(H ₂ O) ₂]Cl.2H ₂ O	3422,b	1695,s	1660,m	1605,m	1129,m	874,w	501,w	470,w

s= sharp, m= medium, w= weak, b= broad, sh=shoulder

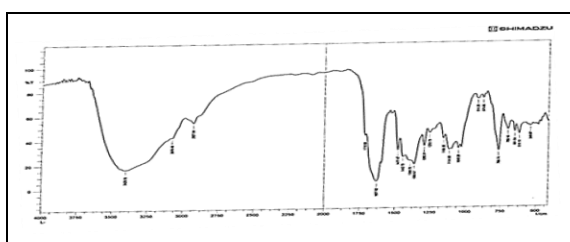


Figure 4. FT-IR spectrum of ONMILA.

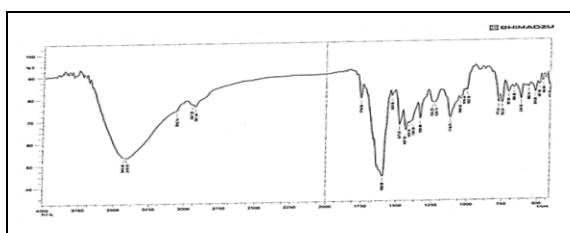


Figure 5. FT-IR spectrum of Ni(II) complex with ONMILA.

3.5 Electronic spectra

Table 3 lists the electronic absorption bands. Uv-Visible spectrum of ONMILA revealed three absorptions as shown in Figure 6 - two at (261 nm, 38341 cm⁻¹) and the other at (334 nm, 29940 cm⁻¹) due to the $\pi-\pi^*$, and one at (411 nm, 24330 cm⁻¹) attributable to the $n-\pi^*$ transition (Ahmed et al., 2015). Two bands that emerged at 825 nm, 12121 cm⁻¹ in Figure 7 electronic spectra for the Co-complex in ethanol solution were ascribed to the ${}^4T_{1g} \rightarrow {}^4A_{2g}$ (ν_2) and ${}^4T_{1g} \rightarrow {}^4T_{1g(p)}$ (ν_3) transitions of octahedral geometry (Ahmed et al., 2015). Using the diagram of Tanaba-Sugano for the d⁷ arrangement of octahedral geometry, the $B_{complex}$ (607.14) value as well as the position of ν_1 (9621 cm⁻¹) were computed using the ratio of (ν_3)/(ν_2) (1.35). (Nisah et al., 2021). The covalent nature is shown by the value of β (0.62). Three bands can be seen in the visible part of the Ni(II) complex's spectrum at (425 nm, 23512 cm⁻¹) ${}^3A_{2g} \rightarrow {}^3T_{1g(p)}$ (ν_3), (702 nm, 14226cm⁻¹) ${}^3A_{2g} \rightarrow {}^3T_{1g(f)}$ (ν_2), the final one at (9183 cm⁻¹) ${}^3A_{2g} \rightarrow {}^3T_{2g(f)}$ (ν_1). On the Tanaba-Sugano diagram, the ratio of ν_2/ν_1 1.54, was used for the d⁸ octahedral complexes (Ahmed et al., 2016), $B_{complex}$ (694.7), and β (0.67). Broadband at (760 nm, 13157 cm⁻¹) in the Cu(II) complex spectrum was due to the ${}^2E_g \rightarrow {}^2T_{2g}$ transition, which is related to the Jahn-Teller distortion

of the octahedral geometry (O'Toole et al., 2019). There was no visible bands in the Cd(II) and Zn(II) complexes spectra as described in Figures 8, 9; instead, there were only bands associated with charge transfer transitions (348 nm, 28735 cm⁻¹) Zn(II) (321 nm, 31152 cm⁻¹) When contrasted were free ligand, there was a presence of two bands at (237 nm, 42194 cm⁻¹) and at (263 nm, 38022 cm⁻¹) (Mutlu et al., 2020).

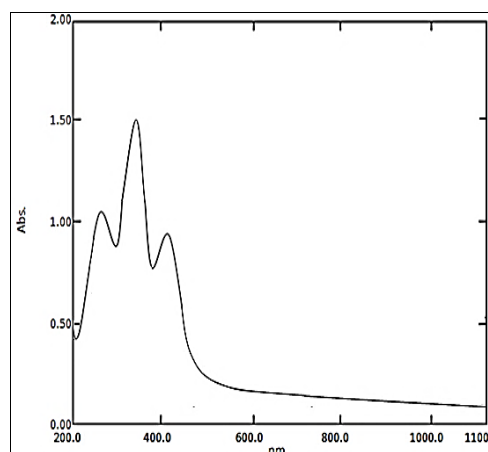


Figure 6. Electronic spectrum of ONMILA.

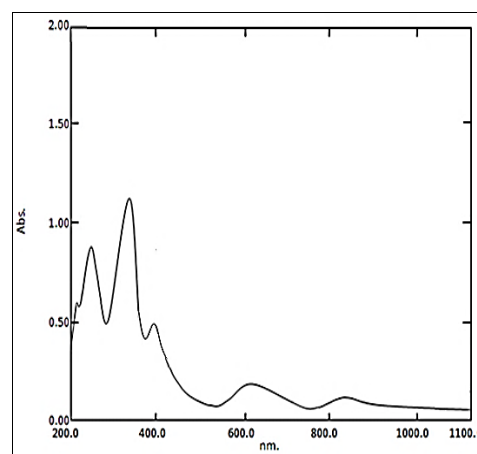


Figure 7. Electronic spectrum of ONMILA-Co(II) complex.

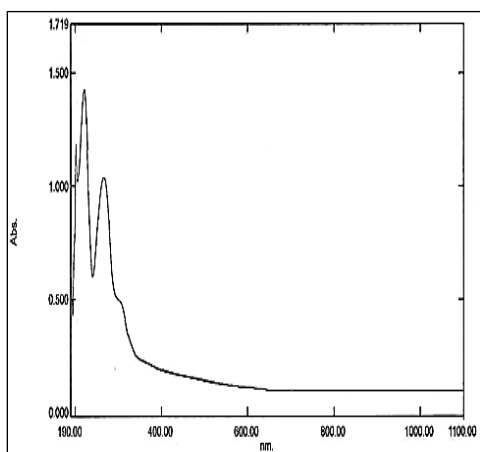


Figure 8. Electronic spectrum of ONMILA-Zn(II) Complex

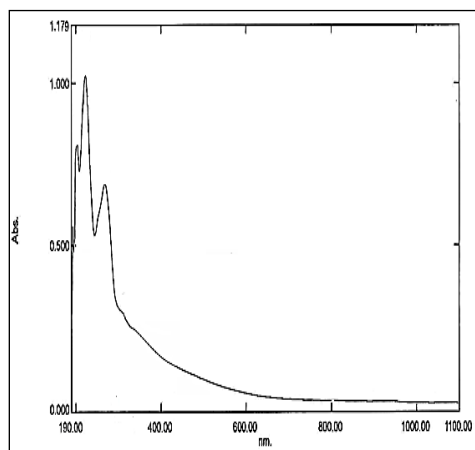


Figure 9. Electronic spectrum of ONMILA-Cd(II) Zn(II) complex.

3.6 Magnetic measurements

The magnetic moment's values possessed by the complexes were used in order to ascertain the coordination of the metal ion. Due to the inherent orbital angular momentum, there is always a big orbital contribution in the ground state, and the effective magnetic moment at ambient temperature was between (4.7 and 5.2 B.M.). The magnetic moment magnitude of the present complex, which was 4.83 B.M., suggested that the Co(II) complex in its high-spin form has the structure of an octahedron (Ahmed et al., 2016). The orbital contribution magnitude is what determines the greater range of values of magnetic moment (2.9-3.4 B.M.) for the complex of a high-spin Ni(II). The magnetic moment obtained in this investigation, 3.16, was in the expected range like octahedral Ni(II) ions (Ahmed et al., 2016). The Cu(II) complex magnetic moment was 1.76 B.M., and it was rather near to the spin value expected for one unpaired electron at 1.73 B.M.. As a result, the Cu(II) complex has the structure of an octahedron (Mutlu et al., 2020; Padmaningrum et al., 2022). According to Sanchez-Lara et al., 2021, the diamagnetic magnetic moments values of the metal complexes Zn(II) and Cd(II) are equal to those of the (d¹⁰) configuration. Table 3 lists the magnetic moments that have been measured.

3.7 Conductivity measurements

All soluble complexes were dissolved in ethanol solvent (1x10⁻³ M) at room temperature and they showed molar conductivity values of (38.07 – 12.95) S.cm². mol⁻¹. The complexes of Co(II), Cu(II) and Ni(II) had low conductivity and a non-ionic structure (Ahmed et al., 2016) while the conductivity tests of Cd(II) and Zn(II) revealed them as electrolytes (Yoe & Jones, 1944). Table 3 shows the conductivity values.

Table 3. Electronic spectra of ONMILA and its metal complexes, measurements of Racah parameter B', nephelauxetic parameter β, magnetic and conductivity values.

Compound	Band position nm	Band position cm ⁻¹	Assignments	B' cm ⁻¹	β	Λ.M S.cm ² .mol ⁻¹	μ _{eff} B.M	Proposed Structure
ONMILA (L)	261	38341	π→π*	—	—	—	—	—
	334	29940						
	411	24330						
L-Co(II)	610	16393	⁴ T _{1g} → ⁴ T _{1g} (p)	607.14	0.62	15.31	4.83	Octahedral Sp ³ d ²
	825	12121	⁴ T _{1g} → ⁴ A _{2g}					
L-Ni(II)	425	23512	³ A _{2g} → ³ T _{1g} (p)	694.7	0.67	12.95	3.16	Octahedral Sp ³ d ²
	702	14226	³ A _{2g} → ³ T _{1g} (F)					
	1088	9183	³ A _{2g} → ³ T _{2g} (F)					
L-Cu(II)	760	13157	² E _g → ² T _{2g}	—	—	16.02	1.76	Octahedral Sp ³ d ²
L-Zn(II)	348	28735	M-L.C.T	—	—	37.84	Dia.	Octahedral Sp ³ d ²
L-Cd(II)	321	31152	M-L C.T	—	—	39.07	Dia.	Octahedral Sp ³ d ²

C.T. = Ligand Field Charge Transfer

3.8 Proposed Molecular Structures of Prepared Complexes

Earlier studies (Ahmed *et al.*, 2016; Mutlu *et al.*, 2020; Nisah *et al.*, 2021; Padmaningrum *et al.*, 2022) have examined the coordination sites available in the ligand and how they are related to various metal ions. The spectroscopic and analytical results based on the molar ratio, elemental analysis metal contents, magnetic measurements and the molar electrical conductivity measurements, as well as the results of the ¹H-NMR spectrum, the ¹³C-NMR spectrum, the UV-Vis spectra and the infrared (FT-IR) spectra all indicated the complexes had octahedral geometry where the ligand behaved as tridentate coordination process. The oxygen of the carbonyl group and two nitrogen atoms of the 6-methyl-2-pyridylmethyl amine bound the ligand to metal ions in addition to water as aqua and chloro ion resulting in six donated atoms to the metal ions, Figures (10 and 11) describe the geometrical aspects of these compounds. There is a 1:1 mole ratio of M to L in these structures.

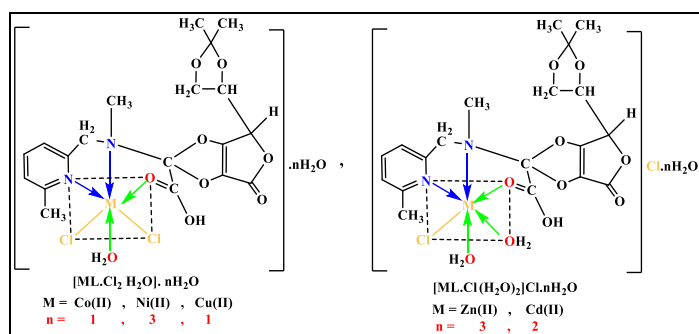


Figure 10. The suggested structure of metals ion complexes with ONMILA.

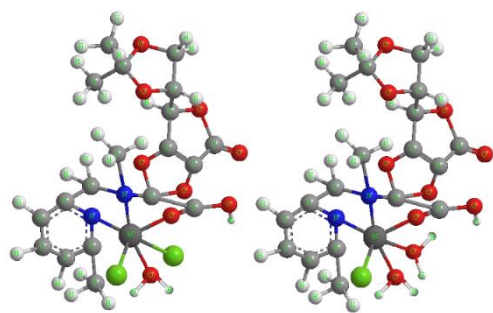


Figure 11. The proposed molecular 3D structure of ONMILA complexes.

3.9 Biological activity

The *in-vitro* growth inhibitory ligand activities and their complexes on Gram (+) bacteria, such as *Staphylococcus aureus* and *Streptococcus pyogenes* and Gram (-) bacteria, namely *Klebsiella pneumonia* and *E-coli*, were investigated using the spotted diffusion method. All of the substances exhibited a significant level of antibacterial activity when tested against the organisms in question. The *Streptococcus pyogenes* and *Staphylococcus aureus* were sensitive to the ligand and its complexes indicating that the biological activity of the complexes ranged from moderate to high. The Cd(II) complex showed higher

antibacterial activity, either *E. coli* or *K. pneumonia*. There was only a slight amount of activity with the ligand and the complexes of Co(II), Cu(II) and Ni(II) while Cu(II) and Zn(II) showed moderate activity. However, the Cd(II) complex showed higher antibacterial activity from either *E. coli* or *K. pneumonia*. The cause of this resistance is the bacteria that live in the colon. These bacteria exist as a single bacillus and have a thick shell that completely encases their cell. The high lipid content of this coating helps to prevent them from entering the cell, in contrast to the bacteria *Staphylococcus aureus* and *Streptococcus pyogenes* which lack this characteristic. They will have a reduced ability to withstand the effects of chemical and antibiotic chemicals that penetrate the inside of the bacterial cell (Shiekhzadeh *et al.*, 2020; Abu-Dief *et al.*, 2020). As a direct consequence of this, the chemical agents that were investigated had a more powerful inhibiting effect. Since the metal ions included inside metal complexes were lipophilic, metal complexes have a higher activity level than free ligands (Omotade *et al.*, 2020). There is also the possibility that these complexes contain antibacterial capabilities which would stop the multiplication of microbes by blocking the active sites of the organisms (Xue *et al.*, 2020). The impermeability of the microorganisms' cells or alterations in the ribosomes of the microbial cells determine the effectiveness of different complexes against the germs that have been tested (Abdel-Rahman *et al.*, 2016). According to Overtone's theory and chelation as described by Tweedy (1964), the metal ion polarity is significantly decreased during the chelation process because of overlaps of the ligand orbital and the slight sharing of the metal ion positive with donor groups. Additionally, the π -electron delocalisation is magnified throughout the chelate sphere resulting in an increase in the lipophilicity of the complex. Through the process of chelation, the core metal atom becomes more lipophilic, which makes it possible for it to traverse the lipid layer of the cell membrane (Pang *et al.*, 2019). Variations in the antibacterial activity are caused by the properties of metal ions as well as the cell membranes of the microorganisms. According to (Hameed *et al.*, 2021), cell wall function inhibition, cell membrane and nucleic acid generation are the four different modes of action that may be attributed to antibacterial medications. These data are presented in Table 4, and the graphical representation of the statistics may be seen in Figure 12.

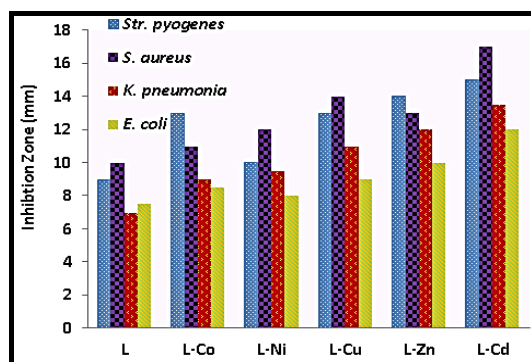


Figure 12. Statistical representation of antibacterial activity for ONMILA and its complexes.

Table 4. Inhibition area (mm) of the bacterial sensitivity (zone inhibition) to the compounds.

Compound	Pathogenic Bacteria			
	<i>Str. pyogenes</i> G(+)	<i>S. aureus</i> G(+)	<i>K. pneumoniae</i> G(-)	<i>E. coli</i> G(-)
Ligand (L)	++	++	+	+
[Co(L)Cl ₂ (H ₂ O)]H ₂ O	+++	++	+	+
[Ni(L)Cl ₂ (H ₂ O)]3H ₂ O	++	+++	+	+
[Cu(L)Cl ₂ (H ₂ O)]H ₂ O	+++	+++	++	+
[Zn(L)Cl(H ₂ O) ₂]Cl.3H ₂ O	++	+++	++	++
[Cd(L)Cl(H ₂ O) ₂]Cl.2H ₂ O	+++	+++	+++	++

Not: (6-9) mm = + (A bit active), (9-12) mm = ++ (Moderate active),

(12-17) mm = +++ (High level of activity)

4. Conclusion

Spectroscopic analyses, such as elemental analysis, metal content, mass spectrum, ¹H, ¹³C-NMR, I.R., and UV-Vis, were used in this research to examine metal ions complexes of the new ligand. Conductivity measurements and magnetic susceptibility testing supported the octahedral geometry of all complexes. There was a 1:1 mole ratio of M to L in these structures. The biological activity of each chemical pointed to its antibacterial properties.

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